NAPA-SOLANO-YOLO-MARIN COUNTY PUBLIC HEALTH LABORATORY

2201 COURAGE DRIVE, MS 9-200 FAIRFIELD, CA 94533 (707) 784-4410 FAX (707) 423-1979









Test	Fungal culture
Test Description	A fungal culture involves screening of tissue samples, swabs, and fluids from miscellaneous body sites for the presence of pathogenic yeasts and molds. Pathogenic species that grow on culture media will be identified to the genus and/or species level. Antibiotic susceptibility testing requires referral and is not typically performed except at the express request of the submitter.
Acceptable specimens and collection	Instructions on specimen collection by specimen type appear below: 1. Cutaneous (skin, nails) a) Cleanse area with an alcohol wipe. b) Obtain skin scrapings or nail clippings in a sterile manner. c) Place specimen in a sterile test tube or container. d) Specimen should remain dry, and kept at room temperature. 2. Hair a) No cleaning of the site necessary. b) From the infected area, use forceps to select at least 10 hairs. c) For hair that is broken off, use a scalpel or a sterile toothbrush to collect scrapings into a sterile container. 3. Upper respiratory tract (ear, nose, nasopharyngeal, mouth) a) Obtain specimen with a sterile, dry swab. b) Place swab in a sterile sheath or test tube. c) Store refrigerated until transported to the laboratory. d) Overnight refrigeration is acceptable. 4. Tissue (fluid or swab biopsies) a) Collect tissue and transport in a sterile screw-cap container with a small amount of nonbacteriostatic saline to prevent drying. b) Do not transport in formalin. 5. Vaginal secretions a) Collect the specimen on a dry swab. b) Place swab in a sterile sheath or container. c) Refrigerate specimen until it can be transported to the laboratory; overnight refrigeration is acceptable. 6. Sputum a) Have patient thoroughly brush teeth and rinse mouth several times with mouthwash, sterile saline or sterile water prior to providing specimen.

	 b) Specimen must be expectorated from deep within the lungs. A first morning specimen is best. Saliva, oral and nasopharyngeal secretions are unsatisfactory. c) Specimen should be collected in a sterile plastic cup with lid. d) Store refrigerated until taken to the laboratory.
	 7. Eye a) Scrape ulcers and suppuration with a Kimura spatula or similar device. b) Inoculate specimen directly onto a blood agar plate or Sabouraud dextrose agar slant, making shallow cuts into the agar. c) Transport promptly to the laboratory. 8. Blood and bone marrow a) Follow routine procedure for collection. b) Place specimen in isolator blood culture collection and mix. 9. Urine a) Obtain a clean-catch specimen (minimum of 10 ml) in a sterile plastic cup with lid. b) Refrigerate specimen until it can be taken to the laboratory. c) Transport to the laboratory within 1–15 hours. Note: Anaerobic transport swabs are not acceptable for fungal sampling.
Specimen storage	Refer to the information above for instructions on specimen storage. In general, normally sterile specimens can be kept at room temperature or at 37°C. Specimens that are potentially contaminated with bacteria should be refrigerated at 4–8°C.
Specimen shipping	Shipping conditions should follow the general collection and storage guidelines above.
CPT code	87118
Test fee	Refer to the posted fee schedule
Result availability (turnaround time)	A preliminary result, describing growth characteristics at week 1, is typically faxed to the submitter within 1 week of specimen receipt. Positive fungal culture results are faxed to the submitter upon first detection and identification over the 4-week culture period. Time to identification after first detection depends upon the bacterial contamination in the specimen, the growth characteristics of the organism, and other factors. Negative specimens require 4 weeks for culturing to be completed. If the specimen is referred to the Microbial Disease Laboratory for further identification, please allow for 1.5 weeks after referral for identification results.